

Molecular Recognition: Hydrogen bonding induced configurational locking of a new Photoresponsive receptor by Dicarboxylic acids

Shyamaprosad Goswami*a, Kumaresh Ghosha, Mintu Halderb

^aDepartment of Chemistry, Bengal Engineering College (Deemed University), Botanic Gardens, Howrah 711 103, India.
^bDepartment of Physical Chemistry, Indian Association for the Cultivation of Science, Jadavpur, Calcutta-700 032, India

Received 8 July 1998; revised 11 December 1998; accepted 21 December 1998

Abstract: A new photoresponsive system 1 has been synthesised and recognition by the cavity of the cisisomer of 1 of dicarboxylic acids of various chain lengths has been studied on irradiation at 310 nm. The cavity of the cis form is found to be selective for adipic acid. © 1999 Published by Elsevier Science Ltd. All rights reserved.

Key words: Molecular recognition, photoresponsive receptor.

Photoresponsive systems are currently attracting much attention because of their many possible applications in diverse fields of science and technology [1-4]. Azo compounds and their derivatives provide such photoresponsive systems which undergo conversion from the trans-to cis-forms on irradiation with UV light ($\lambda = 310$ nm) in solution (although the ground state is mostly populated with the trans-form and reverts to the original trans-form on thermal isomerisation). This photochemical switching leads to two photostationary equilibria based on cis-trans populations [5-8].

Shinkai et al. reported a crown ether containing an aza-photoresponsive receptor [9] which was found to promote photocontrolled extraction and transport of ions across artificial membranes. Photocontrolled reversible association- dissociation phenomena with an ammonio-alkyl lariat in a crown-ring containing an aza- photofunctional system was also studied by Shinkai et al [10]. Azobenzenes have also covalently incorporated into the backbone and also in the chain of many polymers to study the kinetics of cis-trans photoisomerisation of benzene residues in relation to the molecular weight and flexibility of the polymer chain [11-12].

Thus, based on such photochromic behaviour of azobenzenes, we report here a functionalised azo compound 1 having carboxyl binding units at both ends and capable of showing *cis-trans* isomerisation on photoirradiation (though the ground state is mostly *trans*). The cavity of the *cis-isomer* of the photoresponsive system 1 obtained on UV irradiation, selectively recognises adipic acid.

Receptor 1 [13] was synthesised (scheme 1) from azobenzene dicarboxylic acid [8] 2 by coupling

with 2- amino-6-methylpyridine. Compound 1 was isolated in 75% yield.

$$\begin{array}{c}
NO_2 \\
\downarrow \\
NO_2
\end{array}$$

$$\begin{array}{c}
N = N_{\text{tot}} \\
\downarrow \\
CO_2H
\end{array}$$

Scheme 1: i) Glucose, NaOH, ii) PCl₅, 1,2-dichlorobenzene, iii) 2-amino-6-methylpyridine, CH₂Cl₂.

Complexation studies were carried out by NMR titrations as well as by UV experiments using a series of dicarboxylic acids of different chain lengths. As the ground state of the receptor 1 is mostly populated with the trans-form, NMR titration experiments (in CDCl₃ solutions containing 2% d₆-DMSO) give an idea about the complexation ability of the trans-form. The weak binding of dicarboxylic acids with the trans-form of 1 (K_a for glutaric is < 150 M⁻¹; adipic: 1.50 x 10² M⁻¹; suberic: 5.32 x 10² M⁻¹ and sebacic: 1.76 x 10² M⁻¹) is due to the lack of formation of tight complexes with the trans-geometry. In case of glutaric and adipic acids, binding with the trans-form of 1 is significantly weaker and sigmoidal NMR titration curves are observed indicating that other competing modes of binding occur. For long chain suberic and sebacic acids with the trans-form, the break in the titration curves (Δδ vs C_{guest}/ C_{host}) at 0.5 and 1.0 establishes a 2:1 and 1:1 (host: guest) stoichiometry [14] respectively. Our prime objective is to see the interaction of dicarboxylic acids of various chain lengths with the cis-form of 1. Accordingly, UV experiments were performed to study the chain length selectivity by the cavity of the cis-isomer of receptor 1. The cis-isomer of 1 on irradiation at 310 nm, is locked in the presence of aliphatic dicarboxylic acids by the formation of hydrogen bonds, as predicted in complex 2. Formation of this 1:1 complex brings an additional stabilisation of the cisconfiguration over the trans-form of 1 and the extent of stabilisation is strongly dependent on the formation of a tight complex 2 which, in turn, is attributed to the matching of the chain length of dicarboxylic acid with the cavity size of the cis- form of receptor 1.

The positioning of the pyridine amides for binding to the dicarboxylic acids by a suitable spacer is important as shown by the terephthaloyl spacer for adipic acid [15] and the Troger's base spacer for

suberic acid [16]. Here the cavity of the *cis*-form of the receptor 1, where two pyridine amide binding motifs are flanked by a photochemical switching azo unit, is found to be specific for adipic acid.

The absorption spectrum of compound 1 in acetonitrile (c 4.44 x 10^{-5} M) shows a strong band at 335 nm and another centered at 286 nm. On irradiation of the compound with 310 nm light, obtained from a high pressure xenon lamp coupled with a monochromator, the energetically preferred ground state *trans*-form goes to the *cis*-form via a photochemical isomerisation process. The experiments were performed with the receptor 1 in the presence or absence of dicarboxylic acids and the absorption spectrum of 1 was recorded at different time intervals under irradiation. The absorbances at λ_{max} = 335 nm were plotted against time and were found to be fairly linear before the photostationary state is reached.

The cis-form is deduced from the decrease in absorbance (OD) with irradiation time (Fig. 1) (assuming initially the compound is 100% trans) and maximum decrease in OD with time with adipic acid (Fig.2) suggests the tight fitting of the chain into the cavity so that the second step of the following equilibria is shifted more towards the forward direction leading to fast cis-trans isomerisation.

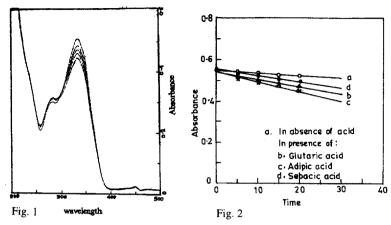
Trans hv Cis dibasic acid Cis (Hydrogen bonded)

Binding studies[†] [17] in CH₃CN at 25° C using UV-VIS spectroscopy show that adipic acid binds to the *trans* form 1 with a binding constant of $5.16 \times 10^3 \text{ M}^{-1}$. Here also the sigmoidal titration curve indicates the several modes of competing binding of the *trans* isomer with adipic acid. In comparison, on irradiation at 310 nm in CH₃CN, the *cis*-form of 1 binds adipic acid with 1:1 stoichiometry by a K_a value of $1.81 \times 10^4 \text{ M}^{-1}$.

In this context, the fully extended conformation of the dicarboxylic acids is more prevalent than the coiled conformation. Dicarboxylic acids with shorter chain lengths that are unable to bridge the two pyridine rings, cannot form the 1:1 complex 2, necessary to lock the cis-isomer of the receptor 1 and this situation is analogous to a free receptor leading to a fast cis-trans isomerisation. Similarly, if the 1:1 complex (as suggested in complex 2) is loose enough, the cis-form, having little stabilisation, will go to the trans-form quickly. If the chain length is larger than the cavity dimension there is also no possibility of formation of 1:1 complex 2. In this regard, formation of 1:1 complexes leading to a dynamic supramolecular structure in solution (i.e. receptor and substrate alternately interlinked) cannot be ruled out. But this situation is also similar to the uncomplexed state having a stabilisation of the trans-form over the cis-form.

The rate of isomerisation of the receptor 1 in the presence of glutaric acid or long chain sebacic acid is almost similar in magnitude but is distinctly modified in the presence of adipic acid. This suggests the formation of a tighter 1:1 complex with adipic rather than glutaric and sebacic acids.

We have thus shown for the first time that the cavity dimension of the cis-isomer of the new azo photoresponsive receptor 1 is selective for adipic acid. Further studies in our laboratory are in progress



to explore this idea in transport phenomena.

Acknowledgement: We thank DST and CSIR, Government of India for financial support and Professor Mihir Chowdhury of Indian Association for the Cultivation of Science, Calcutta for providing the facilities for UV experiments. We also thank the referee for his valuable suggestions.

References:

† UV titrations (CH₃CN): In determining a K_a value with the *trans* form of 1, a solution of the *trans*-form of 1 (8.8 x 10⁻⁵ M) in CH₃CN was used and the absorption was followed at different wavelengths on successive addition of guest solution. The binding constant was determined by Benesi - Hildebrand plot (given in ref. 17). Similarly, in determining a K_a value with the *cis*-form of 1, different sets of stock solutions of *trans*-form with adipic acid of varying composition were prepared. After passing UV light (310 nm) absorption at 335 nm was recorded for each set and the binding constant was determined by the same method.

- [1] Eisenberg SR and Grodzinsky AJ. J. Membr. Sci. 1984;19:173-194.
- [2] Ambilino DB and Stoddart JF. Chem. Rev. 1995;95:2725-2828.
- [3] Dugas H. Bioorganic Chemistry, Springer-Verlag, New York, Inc., 1996.
- [4] Anzai Jl and Osa T. Tetrahedron 1994;50:4039-4067.
- [5] Vogtle F. Supramolecular Chemistry, Wiley; New York, 1993, Ch.7.
- [6]Zimmerman G, Chow LY and Paik UJ. J. Am. Chem. Soc. 1958;80:3528-3531.
- [7] Murakami H, Kawabuchi A, Kotoo K, Kunitake M and Nakashima N. J. Am. Chem. Soc. 1997;119: 7605-7606.
- [8] Ameerunisha S and Zacharias PS. J. Chem. Soc., Perkin Trans 2. 1995;1679-1682.
- [9] Shinkai S. Pure and Appl. Chem. 1987;59:425-430.
- [10] Shinkai S, Yoshida T, Miyazaki K and Manabe O. Bull. Chem. Soc. Jpn. 1987;60:1819-1824.
- [11] Lumarre L and Sung CS. Macromolecules. 1983;16:1729-1736.
- [12] Eisenbach CD. Makromol. Chem. 1979;180:565-571.
- [13] Compound 1 (mp. 268-270°C), M* 450.3 (25.63%), 1 H NMR (200 MHz, CDCl₃) δ : 8.65 (s, 2H, NH), 8.21 (d, 2H, J = 8Hz), 8.12 (d, 4H, J = 8 Hz), 8.06 (d, 4H, J = 8 Hz), 7.68 (t, 2H, J = 8 Hz), 6.97 (d, 2H, J = 8 Hz), 2.50 (s, 6H). 13 C-NMR (50 MHz, CDCl₃): 164.62, 156.87, 154.36, 150.56, 138.80, 136.53, 128.25, 123.30, 119.58, 111.00, 23.85.
- [14] Blanda MT, Horner JH and Newcomb M. J. Org. Chem. 1989;54:4626-4636.
- [15] Tellado FG, Goswami S, Chang SK, Geib SJ and Hamilton AD. J. Am. Chem. Soc. 1990;112:7393-7394.
- [16] Goswami S and Ghosh K. Tetrahedron Lett. 1997;38:4503-4506.
- [17] Colquhoun HM, Goodings EP, Maud JM, Stoddart JF, Wolstenholme JB, Williams DJ. J. Chem. Soc., Perkin Trans II. 1985:607-624.